

VII. READING & INTERPRETING THE RESULTS

■ The upper spot on the ImmunoComb tests for IBD, the middle spot tests for ND and the lower spot tests for IB. Evaluate the results of each disease separately.

■ IBD, ND, IB IgG levels are determined by comparing each specimen's color intensity to the Positive Control (C+). Reading instructions are described in section VI. See illustrations 9 & 10 for details.

■ Specimens with identical or higher color intensity than the Positive Control are considered positive.

■ The Negative Control consists of non-immune sera and should be read as zero (S=0).

■ Non-specific reactions around S1 (i.e., false positives) occurs occasionally due to various reasons and may be associated with the use of certain commercial vaccines. To avoid misinterpretation of non-specific reactions and possible confusion with true low positive results, it is recommended to confirm results by retesting at a one week interval.

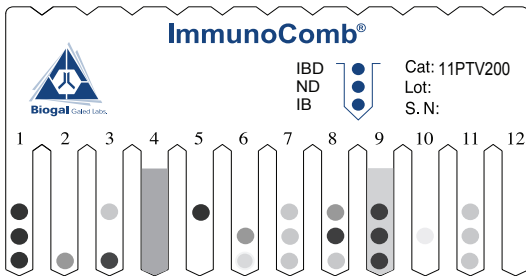
■ A test color darker than S6, indicates either an acute disease or a highly immune flock.

■ Refer to CombScore instructions for score profiling (S) of each specimen antibody level. To determine the immunity profile of your flock use the enclosed CombScore tables (Illustration 11).

■ To assist you interpreting the results, please refer to the example of the developed Comb and the remarks below.

Important: The margin of errors is similar to that of other enzyme Immunoassay kit procedures. Therefore, an error in one color tone will not result in a wrong diagnosis.

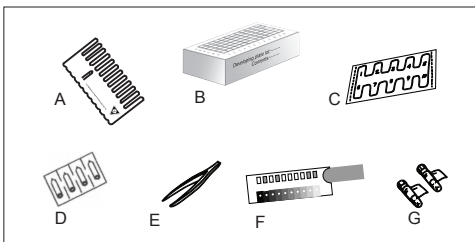
Example of a developed Comb



Tooth No.	Results		
	IB	ND	IBD
1	≥S5 - High Positive	≥S5 - High Positive	≥S5 - High Positive
2	S4 - Positive	Negative	Negative
3	≥S5 - High Positive	Negative	S3 - Positive
4	*Invalid	*Invalid	*Invalid
5	Negative	Negative	≥S5 - High Positive
6	S2 - Inadequate Immunity	S4 Positive	Negative
7	S3 - Positive	S3 - Positive	S3 - Positive
8	S3 - Positive	≥S5 - High Positive	S4 - Positive
9	≥S5 - **Positive	≥S5 - **Positive	≥S5 - **Positive
10	Negative	Negative	Negative
11	S3 - Positive Control	S3 - Positive Control	S3 - Positive Control
12	S0 - Negative Control	S0 - Negative Control	S0 - Negative Control

*High background ** High background with positive reaction

VIII. THE IMMUNOCOMB KIT CONTAINS



A. Three Comb cards, each separately wrapped in an aluminum envelope; **B. Three developing plates** divided into compartments A-F that are subdivided into 12 wells. The plate compartments are pre-filled with reagent solutions; **C. Three specimen papers** with pre-punched disks; **D. Four Blood Lancets**; **E. One pair of disposable tweezers**; **F. One CombScale** color card; **G. One tube of Positive Control** serum and **one tube of Negative Control** serum. **A CombScore sheet** and **an instruction manual** are included.

ImmunoComb

Poultry IBD - ND - IB Antibody Test Kit

50PTV203
INSTRUCTION MANUAL
SUFFICIENT FOR 30 TESTS

I. INTENDED USE

This kit is designed to determine IgG antibody titers to Infectious Bursal Disease (IBD), Newcastle Disease (ND) and Infectious Bronchitis virus (IB; Massachusetts strain) in **chicken and turkey** flocks.

II. WHAT IS THE IMMUNOCOMB ASSAY?

The ImmunoComb is a self-contained portable kit based on a solid phase immunoassay principle. A sensitive test which detects antibody levels in plasma, serum, egg yolk or blood. The ImmunoComb provides results within 38 minutes.

III. HOW DOES THE IMMUNOCOMB WORK?

- The ImmunoComb is a plastic card shaped like a comb, on which purified IBD, ND and IB antigens are attached.
- Samples from chicken or turkey (whole blood saturated paper disks, serum, plasma or egg yolk) are deposited into separate wells in row **A** of the developing plate.
- Positive Control and Negative Control samples, supplied in the kit, should be applied in separate wells in Row **A**.
- Comb is inserted into sample wells so that antibodies from samples bind to the antigens on the Comb's teeth.
- Each plate may be used to test individual or any number of chicken/turkey up to 10, by breaking off the desired number of teeth from the Comb and using the corresponding column of wells in the developing plate. Each run should include Positive Control and Negative Control wells/teeth.
- Non-bound antibodies are washed off in the second row.
- The next row contains an anti-chicken/turkey IgG antibody labeled with an enzyme. While Comb is immersed in this "conjugate", the bound antibodies will be labeled.
- After two washing steps the Comb is inserted into a row where the enzyme reaction takes place. This generates a color change, its intensity indicates the amount of antibodies present in each sample.
- Purple-grey color intensity obtained at each spot should be converted to the antibody levels by using the CombScale.

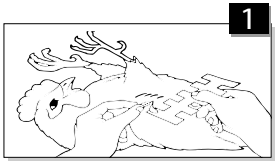
IV. HANDLING & STORAGE

- Store the kit under normal refrigeration: 2° - 8° C (36° - 46° F). **Do not freeze the kit.**
- Before conducting the test, all kit elements and specimens must be at room temperature – preferably for 60 – 120 minutes (or incubate only the developing plate for 22 minutes at 37°C/98.6° F). **Perform assay at room temperature of 20° - 25° C (68° - 77° F).**
- Avoid spillage and cross-contamination of solutions.
- Mix reagents by inverting developing plate several times prior to use.
- Do not mix reagents from different kits or from different rows of the same kit.
- Do not touch teeth of ImmunoComb card.
- When using developing plate, pierce cover of each row by strictly following test procedure instructions. **Do not rip off or remove cover of entire developing plate all at once.**
- The ImmunoComb kit contains inactivated biological material. Kit must be handled and disposed of in accordance with accepted sanitary requirements.

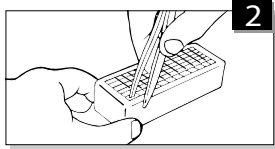
V. STEP-BY-STEP DEVELOPMENT PROCESS

Perform assay at room temperature of 20° - 25° C (68° - 77° F).

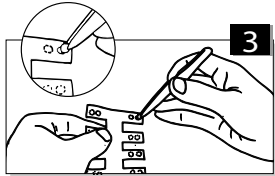
When using blood paper disk



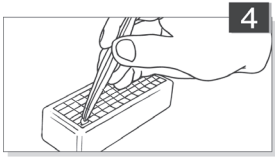
Pierce one of the chick's veins. Take a specimen paper and saturate a pre-punched disk with the blood.



Using the tweezers, slit open the protective aluminum cover of wells in **row A**. One well for each sample



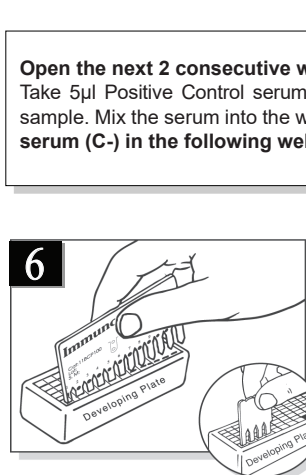
Punch out a disk saturated with blood.



Insert disks into wells of **row A**, dipping into the diluent. **Wait 60 minutes for extraction of antibodies.**

When using egg yolk specimens

Separate the entire egg yolk and wash gently with tap water. Withdraw 1 ml yolk and transfer to a test tube; add 1 ml isotonic saline solution (0.85% NaCl) and mix thoroughly. Deposit 10µl of each diluted yolk specimen into respective wells. Mix by withdrawing and expelling with the pipette several times. Proceed to the next step immediately.

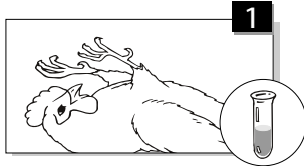


Open the next 2 consecutive wells for control serum.

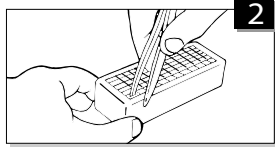
Take 5µl Positive Control serum (C+) and insert into well **A** next to the last sample. Mix the serum into the well. **Do the same with the Negative Control serum (C-) in the following well.**

original envelope for further use. In each further step, open and use only the corresponding wells in the developing plate.

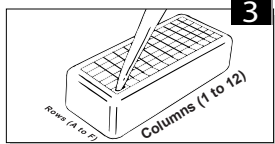
When using serum/plasma



Prepare serum/plasma samples. Apply sample by using either a pipette or a capillary tube.

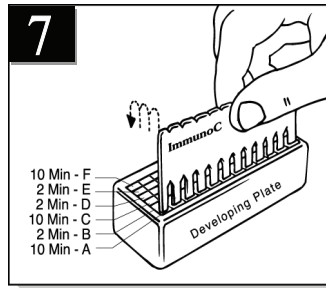


Using the tweezers, slit open the protective aluminum cover of wells in **row A**. One well for each sample

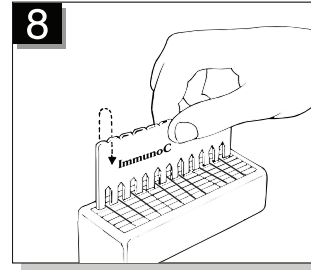


Dispense 5µl sample into each opened well. When using the capillary tubes raise and lower the piston several times to achieve mixing. When using a pipette, mix by depressing the plunger a number of times.

Proceed to step 5.

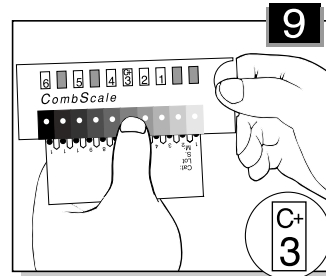


Pierce the cover of wells in **Row B** with the tweezers. Gently shake off excess liquid onto a tissue (follow the same procedure for remaining rows at the end of each step). Insert Comb into wells of **Row B** and incubate for **2 minutes**, shake off and transfer the Comb to **Row C** and incubate for **10 minutes**. Place the Comb in **Row D** for **2 minutes**, **Row E** for **2 minutes**, and **Row F** for **10 minutes**, allowing the color reaction process to develop.



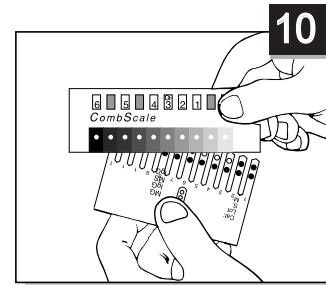
After the Comb has completed the cycle for **Row F**, transfer it back to **Row E**. Incubate in **Row E** for **2 minutes** for color fixation.

VI. READING RESULTS WITH THE COMBSCALE

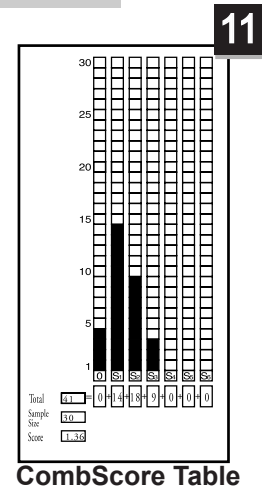


calibrates the C+ to S3, which is the "cut-off" point to which test spots will be compared.

A. When the Comb is completely dry, align it with the calibrated color CombScale provided in the kit. Find the tone of the purple-grey on the CombScale that most closely matches the **Positive Control spot**. Slide the yellow ruler until the C+ mark appears in the window above the color you have found. **Hold the slide in this position during the entire reading.** This step actually



B. Read each of the spots separately: Choose the most suitable color and read the titer in the yellow windows.



CombScore Table



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